Lipoprotein-associated phospholipase A2 (Lp-PLA2)

Lp-PLA2 plays a crucial role in the metabolism of proinflammatory phospholipids and in the generation of pro-atherogenic metabolites, such as lyso-phosphatidylcholine and oxidized free fatty acids. The majority of Lp-PLA2 is found in complexes with LDL, while the remaining portion is bound to HDL. The distribution of Lp-PLA2 over lipoprotein classes correlates with the pathophysiological state of individuals [3]. The association of Lp-PLA2 with lipoproteins is dependent on its glycosylation pattern [4].

Clinical significance of Lp-PLA2

Lp-PLA2 levels have been shown to predict adverse cardiac-related events in both patients with stable coronary artery disease [5] and in a healthy adult population [6]. The increase in Lp-PLA2 levels can predict the development of incident peripheral arterial disease in humans [7]. A meta-analysis that included all prospective studies conducted on Lp-PLA2 with a total of 79,036 patients showed a relationship between the level of Lp-PLA2 and the incidence of coronary artery disease, stroke and cardiovascular mortality in healthy people as well as in patients with a stable vascular disease [8].

Recent guidelines from four major international societies, which include the European Society of Cardiology, the American College of Cardiology, the American Heart Association and the American Society of Endocrinology, have included Lp-PLA2 among the biomarkers through which the measurement is useful for risk stratification of asymptomatic adult patients.

Reagents for the development of quantitative Lp-PLA2 immunoassays

HyTest provides several human Lp-PLA2-specific murine monoclonal antibodies (MAbs) which can be used for the development of immunoassays that enable the detection of Lp-PLA2. In addition, we provide recombinant human Lp-PLA2 protein.

Please note that some Lp-PLA2 assays available are designed to measure the activity of this phospholipase. Our antibodies are suitable for developing a quantitative immunoassay that measures the amount of the biomarker in mass units (ng/ml).

Monoclonal antibodies specific to Lp-PLA2

We provide four well-characterized human Lp-PLA2-specific MAbs for the detection of Lp-PLA2 from plasma samples. The antibodies were developed against a recombinant human Lp-PLA2 expressed in a mammalian cell line.

A quantitative sandwich immunoassay for Lp-PLA2

We recommend two MAb combinations for the development of a sandwich immunoassay to measure Lp-PLA2 in human plasma samples: PL42cc–PL46cc and PL26cc–PL4cc. Also other pairs are possible. All HyTest MAb combinations are capable of detecting native Lp-PLA2 in human serum or plasma.

All pair recommendations are listed in Table 1. Meanwhile, the calibration curve for the combination PL42cc–PL46cc is provided in Figure 1.

Table 1. The most sensitive capture-detection pairs. Data is based on the results obtained using our in-house fluoroimmunoassay.

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<tr>
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<td>PL26cc</td>
<td>PL4cc</td>
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Figure 1. Calibration curve for the MAb combination PL42cc–PL46cc. The capture antibody PL42cc was coated onto the wells of a Costar EIA/RIA plate and the plate was blocked with a buffer that contained 1% casein and 0.05% Tween 20 at room temperature for 15 minutes. Recombinant human Lp-PLA2 (Cat.# 8PL7) and the Eu³⁺-labeled detection MAb PL46cc were diluted in an assay buffer and incubated in coated plate wells for 2.5 hours at 37°C. Note: Due to the lipid binding properties of Lp-PLA2 it has a tendency to non-specifically adsorb onto plastic surfaces. To avoid this, 1% casein was added to the assay buffer to 0.5% (v/v) final concentration.

Figure 2. Dilutional linearity study. Dilutional linearity study of recombinant Lp-PLA2 and native Lp-PLA2 (normal human serum from an apparently healthy volunteer) studied using the MAb combination PL42cc–PL46cc. The protocol is described in the Figure 1 caption. The initial concentration of the recombinant human Lp-PLA2 was 111 ng/ml.

All HyTest antibodies are suitable for labeling with horseradish peroxidase. Figure 3 provides examples of sandwich ELISA assays that utilize HRP-labeled detection antibodies.

Figure 3. The titration curve of recombinant Lp-PLA2 (Cat.# 8PL7) in sandwich ELISA using HRP-labelled antibodies. Plate wells coated with coating antibodies (1 µg per well) were blocked with 1% casein to prevent non-specific adsorption of antigen onto the plate surface. Recombinant Lp-PLA2 was added in indicated concentrations followed by HRP-linked detection antibodies (0.4 µg per well).
Measuring patient samples

In order to conduct preliminary clinical studies, we obtained serum samples from patients diagnosed with AMI at admission (N=13) as well as from apparently healthy volunteers (N=13). Figure 4 shows the box-whisker plots of immunoassays that utilize MAbs PL26cc–PL4cc (A) and PL42cc–PL46cc (B) or a commercially available diagnostic assay (C) (see Figure 4).

HyTest’s MAb pair PL26cc–PL4cc detected native Lp-PLA2 in a manner that was very similar to that of the commercially available ELISA assay. Meanwhile, the PL42cc–PL46cc assay detected native Lp-PLA2 in a slightly different way.

Recombinant human Lp-PLA2

HyTest provides recombinant human Lp-PLA2 (recLp-PLA2) that is expressed in a mammalian cell line. The protein contains 6×His tag on its C-terminus linked with a GG spacer. The calculated molecular mass and isoelectric point of recLp-PLA2 are 49,000 Da and 7.1, respectively. RecLp-PLA2 contains N-linked glycans like its native counterpart (data not shown).

RecLp-PLA2 is purified from the conditioned media of the mammalian cell line with several chromatographic procedures. Isolated recLp-PLA2 is substantially free of contaminants and has purity >75% as determined by densitography following SDS-gel electrophoresis in reducing conditions (data not shown).

Recombinant Lp-PLA2 associates with plasma fractions like endogenous Lp-PLA2

In gel-filtration, the immunoreactivity of native Lp-PLA2 is distributed over the entire elution profile that forms three distinct peaks. Elution volumes of the peaks are in good accordance with those for VLDL, LDL and HDL, i.e. lipoprotein particles with which the native Lp-PLA2 forms complexes [9]. RecLp-PLA2 spiked in normal human serum showed that the elution profile of the recombinant protein was similar to that of endogenous Lp-PLA2 (see Figure 5). The peaks coincide with those of the native protein which indicates that recLp-PLA2 binds to the same moieties in the serum as native Lp-PLA2.

Figure 5. Gel-filtration studies of the association of recombinant human Lp-PLA2 with lipoproteins in normal human serum. 150 µl of normal human serum or 150 µl of serum spiked with 3 µg of recLp-PLA2 were applied onto a Superose 6 gel-filtration column. Immunoreactivity in fractions was determined by a fluoroimmunoassay with the MAb combination PL42cc–PL46cc. RecLp-PLA2 appears to be binding to the same lipoproteins as endogenous Lp-PLA2 when added to native serum.

Recovery of recombinant Lp-PLA2 in human serum

When recLp-PLA2 is spiked in normal human serum, the resulting signal represents a summary of a signal with recLp-PLA2 alone in an assay buffer and native Lp-PLA2 in serum over a wide concentration range (see Figure 6).

Stability of recombinant human Lp-PLA2

The recombinant human Lp-PLA2 is sold as a lyophilized product. However, we investigated how a protein preparation in a storage buffer retained its immunoreactivity when stored at different temperatures (see Figure 7). Based on the results, recLp-PLA2 retained approximately 80% of its immunoreactivity for at least two weeks when stored at room temperature. Meanwhile, when stored at 37°C, 50% of the immunoreactivity was lost after four days.

Ordering information

MONOCLONAL ANTIBODIES

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ANTIGEN

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Figure 6. Recovery studies of recLp-PLA2 in normal human serum with the MAb combination PL42cc–PL46cc. recLp-PLA2 was spiked into normal human serum to concentrations indicated. The same concentrations of recLp-PLA2 were used for measurement in an assay buffer. Serum dilutions were prepared with the assay buffer and used for the native Lp-PLA2 immunoreactivity measurement.

Figure 7. Short-term temperature stability of recLp-PLA2. Aliquots of recLp-PLA2 solution were incubated at room temperature or at 37°C for two weeks. At indicated time points, aliquots were diluted to an assay buffer and recLp-PLA2 immunoreactivity was measured using the MAb combination PL42cc–PL46cc.

The immunoreactivity of recLp-PLA2 measured with the MAb combination PL42cc–PL46cc does not change significantly over 15 freeze-thaw cycles (see Figure 8).